

## SERVA DNA Standard pUC 19 x MspI, lyophilized

Cat. No.: 39304.01

Qty.: 1 x 50 µg

Fragment sizes (in Bp): 501, 489, 404, 331, 242, 190, 147, 111, 110, 67, 34 (2x), 26

### How to dissolve the lyophilized DNA fragments:

Depending on the usage of the marker dissolve the DNA fragments in 1 x sample buffer (1 ml steril filtered sample buffer is supplied with each marker, for composition please see below) or, e.g. for subsequent labelling of DNA fragments, in TE buffer (10 mM Tris/HCl pH 7.5 and 1 mM EDTA). Add the volume of buffer needed for your application to the lyophilized DNA and incubate for 15 min at room temperature.

Buffer volume	50 µl	100 µl	250 µl	500 µl	1000 µl
µg DNA in 1 µl	1	0.5	0.2	0.1	0.05
µg DNA in 5 µl	5	2.5	1	0.5	0.25
µg DNA in 10 µl	10	5	2	1	0.5

### Sample application:

for UV detection after EtBr. staining:

use 0.5 - 1.0 µg DNA fragments per lane

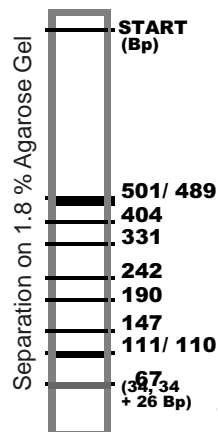
for detection using signal integrating systems after EtBr. staining:

use 0.1 - 0.5 µg per lane

### Storage:

Store lyophilized DNA fragments at -20 °C, store resuspended DNA fragments at -20 °C; avoid repeated thaw/freeze cycles (> 10fold), if needed, aliquot the marker fragments.

1 x sample buffer (TE/G-buffer)	
Tris/HCl pH 7.5	10 mM
Sodium-acetate	5 mM
EDTA	2 mM
Glycerol	10 % (w/v)
Bromophenolblue	0.03 % (w/v)
Xylene Cyanol FF	0.02 % (w/v)



## SERVA DNA Standard $\lambda$ x BstEII, lyophilized

Cat. No.: 39301.01

Qty.: 2 x 50  $\mu$ g

Fragment sizes (in Bp): 8.543, 7.242, 6.369, 5.687, 4.822, 4.324, 3.675, 2.323, 1.929, 1.371, 1.264, 702, 224, 117

### How to dissolve the lyophilized DNA fragments:

Depending on the usage of the marker dissolve the DNA fragments in 1 x sample buffer (1 ml steril filtered sample buffer is supplied with each marker, for composition please see below) or, e.g. for subsequent labelling of DNA fragments, in TE buffer (10 mM Tris/HCl pH 7.5 and 1 mM EDTA). Add the volume of buffer needed for your application to the lyophilized DNA and incubate for 15 min at room temperature.

Buffer volume	50 $\mu$ l	100 $\mu$ l	250 $\mu$ l	500 $\mu$ l	1000 $\mu$ l
$\mu$ g DNA in 1 $\mu$ l	1	0.5	0.2	0.1	0.05
$\mu$ g DNA in 5 $\mu$ l	5	2.5	1	0.5	0.25
$\mu$ g DNA in 10 $\mu$ l	10	5	2	1	0.5

### Sample application:

for UV detection after EtBr. staining:

use 0.5 - 1.0  $\mu$ g DNA fragments per lane

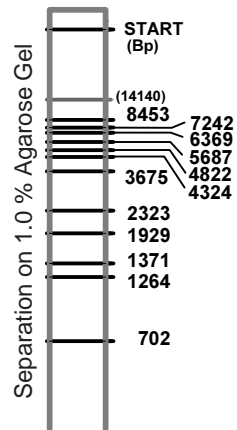
for detection using signal integrating systems after EtBr. staining:

use 0.1 - 0.5  $\mu$ g per lane

### Storage:

Store lyophilized DNA fragments at -20 °C, store resuspended DNA fragments at -20 °C; avoid repeated thaw/freeze cycles (> 10fold), if needed, aliquot the marker fragments.

1 x sample buffer (TE/G-buffer)	
Tris/HCl pH 7.5	10 mM
Sodium-acetate	5 mM
EDTA	2 mM
Glycerol	10 % (w/v)
Bromophenolblue	0.03 % (w/v)
Xylene Cyanol FF	0.02 % (w/v)



## SERVA DNA Standard pBR 328 Mix, lyophilized

Cat. No.: 39302.01

Qty.: 1 x 50 µg

**Fragment sizes (in Bp): 2.176, 1.766, 1.230, 1.033, 653, 517, 453, 394, 298, 234, 220, 154**

### How to dissolve the lyophilized DNA fragments:

Depending on the usage of the marker dissolve the DNA fragments in 1 x sample buffer (1 ml steril filtered sample buffer is supplied with each marker, for composition please see below) or, e.g. for subsequent labelling of DNA fragments, in TE buffer (10 mM Tris/HCl pH 7.5 and 1 mM EDTA). Add the volume of buffer needed for your application to the lyophilized DNA and incubate for 15 min at room temperature.

Buffer volume	50 µl	100 µl	250 µl	500 µl	1000 µl
µg DNA in 1 µl	1	0.5	0.2	0.1	0.05
µg DNA in 5 µl	5	2.5	1	0.5	0.25
µg DNA in 10 µl	10	5	2	1	0.5

### Sample application:

for UV detection after EtBr. staining:

use 0.5 - 1.0 µg DNA fragments per lane

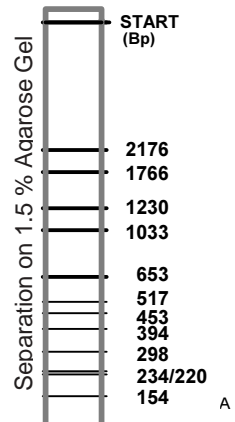
for detection using signal integrating systems after EtBr. staining:

use 0.1 - 0.5 µg per lane

### Storage:

Store lyophilized DNA fragments at -20 °C, store resuspended DNA fragments at -20 °C; avoid repeated thaw/freeze cycles (> 10fold), if needed, aliquot the marker fragments.

1 x sample buffer (TE/G-buffer)	
Tris/HCl pH 7.5	10 mM
Sodium-acetate	5 mM
EDTA	2 mM
Glycerol	10 % (w/v)
Bromophenolblue	0.03 % (w/v)
Xylene Cyanol FF	0.02 % (w/v)



## SERVA DNA Standard pBR 322 x HaellI, lyophilized

Cat. No.: 39303.01

Qty.: 1 x 50 µg

Fragment sizes (in Bp): 587, 540, 502, 458, 434, 267, 234, 213, 192, 184, 124, 123, 104, 89, 80, 64, 57, 51, 21, 18, 11, 8

### How to dissolve the lyophilized DNA fragments:

Depending on the usage of the marker dissolve the DNA fragments in 1 x sample buffer (1 ml steril filtered sample buffer is supplied with each marker, for composition please see below) or, e.g. for subsequent labelling of DNA fragments, in TE buffer (10 mM Tris/HCl pH 7.5 and 1 mM EDTA). Add the volume of buffer needed for your application to the lyophilized DNA and incubate for 15 min at room temperature.

Buffer volume	50 µl	100 µl	250 µl	500 µl	1000 µl
µg DNA in 1 µl	1	0.5	0.2	0.1	0.05
µg DNA in 5 µl	5	2.5	1	0.5	0.25
µg DNA in 10 µl	10	5	2	1	0.5

### Sample application:

for UV detection after EtBr. staining:

use 0.5 - 1.0 µg DNA fragments per lane

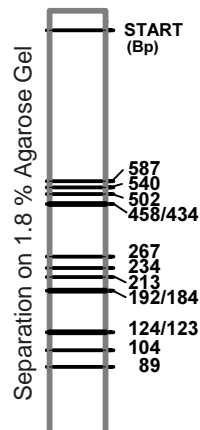
for detection using signal integrating systems after EtBr. staining:

use 0.1 - 0.5 µg per lane

### Storage:

Store lyophilized DNA fragments at -20 °C, store resuspended DNA fragments at -20 °C; avoid repeated thaw/freeze cycles (> 10fold), if needed, aliquot the marker fragments.

1 x sample buffer (TE/G-buffer)	
Tris/HCl pH 7.5	10 mM
Sodium-acetate	5 mM
EDTA	2 mM
Glycerol	10 % (w/v)
Bromophenolblue	0.03 % (w/v)
Xylene Cyanol FF	0.02 % (w/v)



## SERVA DNA Standard 100 Bp DNA Ladder equimolar, lyophilized

Cat. No.: 39311.01 Qty.: 1 x 50 µg

Fragment sizes (in Bp): 1.000, 900, 800, 700, 600, 500 (2x),  
400, 300, 200, 100

### How to dissolve the lyophilized DNA fragments:

Depending on the usage of the marker dissolve the DNA fragments in 1 x sample buffer (1 ml steril filtered sample buffer is supplied with each marker, for composition please see below) or, e.g. for subsequent labelling of DNA fragments, in TE buffer (10 mM Tris/HCl pH 7.5 and 1 mM EDTA). Add the volume of buffer needed for your application to the lyophilized DNA and incubate for 15 min at room temperature.

Buffer volume	50 µl	100 µl	250 µl	500 µl	1000 µl
µg DNA in 1 µl	1	0.5	0.2	0.1	0.05
µg DNA in 5 µl	5	2.5	1	0.5	0.25
µg DNA in 10 µl	10	5	2	1	0.5

### Sample application:

for UV detection after EtBr. staining:

use 0.3 - 0.8 µg DNA fragments per lane

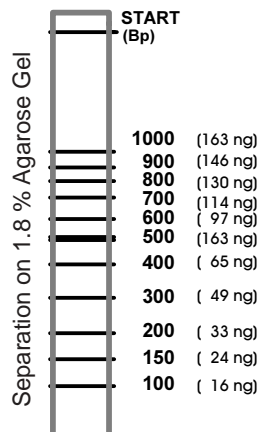
for detection using signal integrating systems after EtBr. staining:

use 0.05 - 0.3 µg per lane

### Storage:

Store lyophilized DNA fragments at -20 °C,  
store resuspended DNA fragments at -20 °C;  
avoid repeated thaw/freeze cycles (> 10fold),  
if needed, aliquot the marker fragments.

1 x sample buffer (TE/G-buffer)	
Tris/HCl pH 7.5	10 mM
Sodium-acetate	5 mM
EDTA	2 mM
Glycerol	10 % (w/v)
Bromophenolblue	0.03 % (w/v)
Xylene Cyanol FF	0.02 % (w/v)



## SERVA DNA Standard 100 Bp DNA Ladder extended, lyophilized

Cat. No.: 39312.01

Qty.: 1 x 50 µg

Fragment sizes (in Bp): 5.000, 4.000, 3.000, 2.500, 2.000,  
1.500, 1.000, 900, 800, 700, 600, 500 (2x), 400, 300, 200, 100

### How to dissolve the lyophilized DNA fragments:

Depending on the usage of the marker dissolve the DNA fragments in 1 x sample buffer (1 ml steril filtered sample buffer is supplied with each marker, for composition please see below) or, e.g. for subsequent labelling of DNA fragments, in TE buffer (10 mM Tris/HCl pH 7.5 and 1 mM EDTA). Add the volume of buffer needed for your application to the lyophilized DNA and incubate for 15 min at room temperature.

Buffer volume	50 µl	100 µl	250 µl	500 µl	1000 µl
µg DNA in 1 µl	1	0.5	0.2	0.1	0.05
µg DNA in 5 µl	5	2.5	1	0.5	0.25
µg DNA in 10 µl	10	5	2	1	0.5

### Sample application:

for UV detection after EtBr. staining:

use 0.5 - 0.8 µg DNA fragments per lane

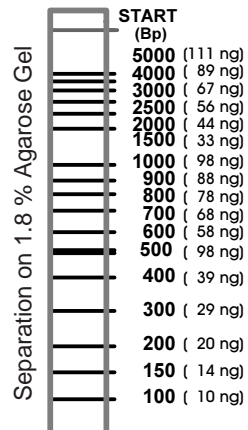
for detection using signal integrating systems after EtBr. staining:

use 0.2 - 0.5 µg per lane

### Storage:

Store lyophilized DNA fragments at -20 °C,  
store resuspended DNA fragments at -20 °C;  
avoid repeated thaw/freeze cycles (> 10fold),  
if needed, aliquot the marker fragments.

1 x sample buffer (TE/G-buffer)	
Tris/HCl pH 7.5	10 mM
Sodium-acetate	5 mM
EDTA	2 mM
Glycerol	10 % (w/v)
Bromophenolblue	0.03 % (w/v)
Xylene caynol FF	0.02 % (w/v)



## SERVA DNA Standard 100 Bp DNA Ladder equalized, lyophilized

Cat. No.: 39313.01 Qty.: 1 x 20 µg

Fragment sizes (in Bp): 1.000, 900, 800, 700, 600, 500 (2x),  
400, 300, 200, 100

### How to dissolve the lyophilized DNA fragments:

Depending on the usage of the marker dissolve the DNA fragments in 1 x sample buffer (1 ml steril filtered sample buffer is supplied with each marker, for composition please see below) or, e.g. for subsequent labelling of DNA fragments, in TE buffer (10 mM Tris/HCl pH 7.5 and 1 mM EDTA). Add the volume of buffer needed for your application to the lyophilized DNA and incubate for 15 min at room temperature.

Buffer volume	50 µl	100 µl	250 µl	500 µl	1000 µl
µg DNA in 1 µl	0.4	0.2	0.08	0.04	0.02
µg DNA in 5 µl	2	1	0.4	0.2	0.1
µg DNA in 10 µl	4	2	0.8	0.4	0.2

### Sample application:

for UV detection after EtBr. staining:

use 0.2 - 0.5 µg DNA fragments per lane

for detection using signal integrating systems after EtBr. staining:

use 0.05 - 0.2 µg per lane

### Storage:

Store lyophilized DNA fragments at -20 °C,  
store resuspended DNA fragments at -20 °C;  
avoid repeated thaw/freezing cycles (> 10fold),  
if needed, aliquot the marker fragments.

1 x sample buffer (TE/G-buffer)	
Tris/HCl pH 7.5	10 mM
Sodium-acetate	5 mM
EDTA	2 mM
Glycerol	10 % (w/v)
Bromophenolblue	0.03 % (w/v)
Xylene Cyanol FF	0.02 % (w/v)

